

# Sperm MorfoStain™

## Intended Use: Summary and Explanation

Sperm MorfoStain™ is a classical Romanovsky type stain. It is a one step stain optimised for assessment of sperm morphology, which is one of the basic tools used in semen analysis. The technique is based on the principle that the sperm will be stained in a darker colour (blue) and the background will be lighter and therefore the shape, size and integrity of the sperm can easily be seen in oil immersion under a microscope.

## Components

Methanol  
Eosine Y  
Methylene B  
May Grunwald  
Giemsa  
Azur B

## Storage and Stability

Store at 10-25°C. Not in a refrigerator. Under these conditions Sperm MorfoStain™ has a shelf-life of 24 months. The expiry date is shown on the bottle.

## Precautions and Warnings

- Sperm MofoStain™ is highly flammable
- Always handle the product in a safety cabinet
- Inhalation: Remove from exposure. If discomfort persists. Obtain medical attention
- Skin contact: Wash of with abundance of water. Remove contaminated clothing
- Eye contact: Flush thoroughly with physiological saline for 15 min. Obtain medical attention

- Ingestion: Rinse mouth with water and give water to drink. Obtain medical attention. Do not induce vomiting
- Do not use any solution which shows evidence of bacterial contamination or if stopper accidentally comes in contact with unsterile surfaces
- MSDS is available from the distributor or manufacturer (see nidacon.com)

## Ordering Information

**Volume**  
250mL

**Article No.**  
SMS-250



[www.nidacon.com](http://www.nidacon.com)

**For further technical information or assistance**, please contact your distributor or the manufacturer.



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### Reagents and Equipment

- Sperm MorfoStain™
- Coplin jar or similar
- Glas microscope slide holder
- Microscope slides
- Light microscope (400-1000x magnification)
- Pipette

### Procedure for staining with Sperm MorfoStain™

Use pencil to mark your samples since the stain will remove permanent marker.

1. Let sample liquify for 30 minutes before preparation
2. Make a semen smear on a glas microscope slide by conventional method or method shown in steps 3-4
3. Transfer a 20µl droplet of semen onto a labelled microscope slide with a pipette, making a string/line of fluid in the middle of the slide
4. Cover this slide with a second microscope slide and, when the droplet is evenly spread between the two slides, pull them apart from each other horizontally. This method will give you two good slides
5. Let smears air dry
6. Dip the dry smears into the staining solution for max 10 sec
7. Rinse in double distilled water, changing the water 3 times. Let slides air dry lying flat
8. Mount the slides with coverslips and DPX or equivalent mounting fluid. Let dry completely. Examine using a bright-field 100x objective under oil immersion
9. Count at least 200 sperm, and classify them as normal or abnormal according to the 2002 NAFA and ESHRE-SIGA Manual on Basic Semen Analysis.

